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Evidence that Endogenous Vasoactive Intestinal Peptide (VIP) is Involved in the Regulation of Rat Pituitary-adrenocortical Function: in vivo Studies with a VIP Antagonist

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Abstract—The effect of a subcutaneous bolus injection of 2 µg |Ac,Tyr¹,D-Phe²|-GRF(1–29) amide, a specific VIP antagonist (VIP-A), on the hypothalamo-pituitary-adrenocortical (HPA) axis were investigated in both normal and ether- or cold-stressed rats. Blood concentrations of ACTH, aldosterone (ALDO) and corticosterone (B) were measured by specific RIA 1, 2 or 4 h after VIP-A injection. VIP-A administration to normal rats strikingly lowered the plasma concentration of ALDO, without significantly affecting those of ACTH and B. Ether and cold stresses notably raised the blood levels of ACTH, ALDO and B, and these rises lasted unchanged until 4 h. VIP-A did not affect the response of HPA axis to ether stress, but provoked a marked depression of that to cold stress. In light of these findings the following conclusions can be drawn: (i) endogenous VIP does not regulate HPA-axis function under basal conditions, but it plays a pivotal role in the mechanisms involved in the activation of HPA axis induced by cold exposure; and (ii) endogenous VIP exerts a tonic stimulatory action on ALDO secretion, probably by acting directly on the adrenal zona glomerulosa.

Introduction

VIP is a strongly basic octacosapeptide widely distributed in the central and peripheral nervous systems, that at present is recognized as a

neurotransmitter involved in neuroendocrine regulation.^{1–3}

VIP is present in all components of the hypothalamo-pituitary-adrenal (HPA) axis. In the hypothalamus, the highest concentration of this neuropeptide is found in the suprachiasmatic nucleus, while in the cells of the parvocellular subdivision of the paraventricular nucleus VIP is co-

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localized with CRH.⁴⁻⁶ These neurons project fibers to the external zone of the median eminence, where VIP can be released into the portal circulation.^{7,8} VIP immunoreactivity and VIP mRNA have been demonstrated in the anterior pituitary, and in an in-vitro perfusion system ACTH was found to cause an appreciable release of pituitary VIP.^{2,9-12} Adrenal zona medullaris contains a small number of VIP-positive cells, and an abundant network of VIP-ergic fibers is present in zona medullaris, as well as in the adrenal capsule and adjacent zona glomerulosa.¹³⁻¹⁸

The above described localization of VIP strongly suggests its involvement in the regulation of HPA-axis function. Indeed, various investigations confirm a role of VIP in the regulation of ACTH and adrenocortical-steroid release.^{2,19} The present study was designed to ascertain whether endogenous VIP plays a role in the functional regulation of rat HPA axis under basal and stress conditions. To achieve this goal, a specific competitive antagonist of VIP (VIP-A), the |Ac-Tyr¹,D-Phe²|-GRF(1-29) amide,²⁰ was employed.

Methods

Adult female Wistar rats weighing 190–210 g were kept under a 10:14 h light-dark cycle (illumination onset at 8:00 a.m.) at $22 \pm 1^\circ\text{C}$. They were divided into 3 groups, two of which were stressed by ether inhalation or cold exposure. *Ether stress*: rats were individually placed for 2 min in a 10 l-glass jar, in which 10 ml of ethyl ether had been previously poured, and employed 10 min later.²¹ *Cold stress*: rats were placed in a wire cage for 20 min at 4°C , and immediately employed. Each of the 3 groups of rats was divided into two subgroups; one subgroup was subcutaneously administered VIP-A (Peninsula, Merseyside, UK) dissolved in isotonic saline (2 $\mu\text{g}/\text{rat}$), the other subgroup received vehicle only and served as a control. All the injections were made at 9:00 a.m., and rats were decapitated at times 0 (no injection), 1, 2 or 4 h after the injection (6 rats for each time point). The trunk blood of each rat was collected in the presence of NaF (1 $\text{mg}\cdot\text{ml}^{-1}$), plasma was separated and stored at -20°C .

ACTH was extracted from plasma, and its con-

centration determined by RIA: ACTHK-RIA kit (Cis Bio International, Gif-sur-Yvette, France). Sensitivity: 10 $\text{pg}\cdot\text{ml}^{-1}$. Cross-reactivity: ACTH(1–24), 100%; α -MSH and β -lipotropin, 0.1%; other pituitary hormones, less than 0.001%. Intra-assay and inter-assay variations: 6% and 9%, respectively. Aldosterone (ALDO) and corticosterone (B) were extracted from plasma, and their concentrations measured by RIA, using |1,2,6,7-³H|-ALDO and |1 α ,2 α (n)-³H|-B (Amersham, UK; S.A.: 1.96 $\text{TBq}\cdot\text{mmol}^{-1}$) and antisera developed in the rabbit (Sigma, St. Louis, MO, USA). *ALDO-RIA*: sensitivity: 5 $\text{pg}\cdot\text{ml}^{-1}$. Cross-reactivity: ALDO, 100%; 17-iso-ALDO and other steroids, less than 0.1%. Intra-assay and inter-assay variations: 5% and 7%, respectively. *B-RIA*: sensitivity: 50 $\text{pg}\cdot\text{ml}^{-1}$. Cross-reactivity: B and cortisol, 100%; 11-deoxycorticosterone and progesterone, 2%; other steroids, less than 0.001%. Intra-assay and inter-assay variations: 7% and 9%, respectively. Blood glucose concentration was estimated by a commercial kit (Glucose Hexokinase; Abbott Labs, Chicago, Ill., USA; c.v.: 3%).

Data were expressed as means \pm SE ($n = 6$), and their statistical comparison was done by ANOVA, followed by the Multiple Range test of Duncan.

Results

Non-stressed rats

Vehicle injection resulted in transient significant increases in the plasma concentrations of ACTH (2-fold), ALDO (68%) and B (2.3-fold) at 1 h, followed by return to the baseline at 4 h (Figs 1–3). Glycemia was not affected (Fig. 4).

VIP-A administration did not affect plasma ACTH level at 1 h, though raising it over the respective control value at 2 h (75%) and 4 h (3-fold) (Fig. 1). VIP-A induced a notable drop in ALDO blood level at 1 h (-54%), 2 h (-51%) and 4 h (-43%) (Fig. 2). B blood level and glycemia were not significantly changed by VIP-A (Figs 3 and 4).

Stressed rats

As expected, both ether and cold stresses notably raised the blood levels of ACTH (6.5-fold and 8.5-fold, respectively), ALDO (3.6-fold and 3.7-fold), B (6.7-fold and 8.8-fold) and glucose (12% and

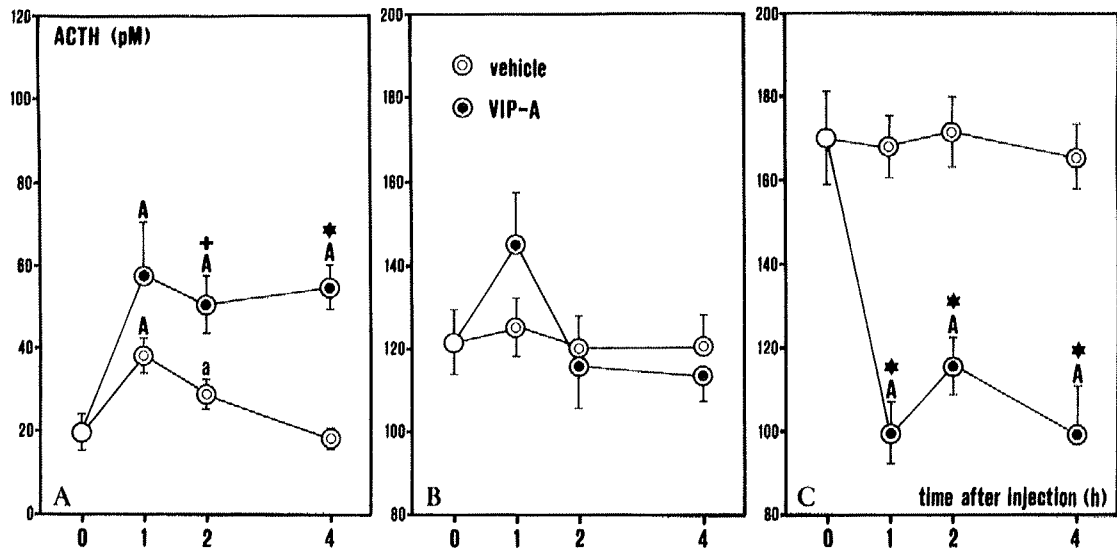


Fig. 1 Effect of VIP-A on ACTH plasma concentration of normal (A), ether-stressed (B) and cold-stressed (C) rats. Data are means \pm SE ($n = 6$). $^aP < 0.05$ and $^AP < 0.01$ from 0-h group; $^+P < 0.05$ and $^*P < 0.01$ from the respective control (vehicle-injected) group.

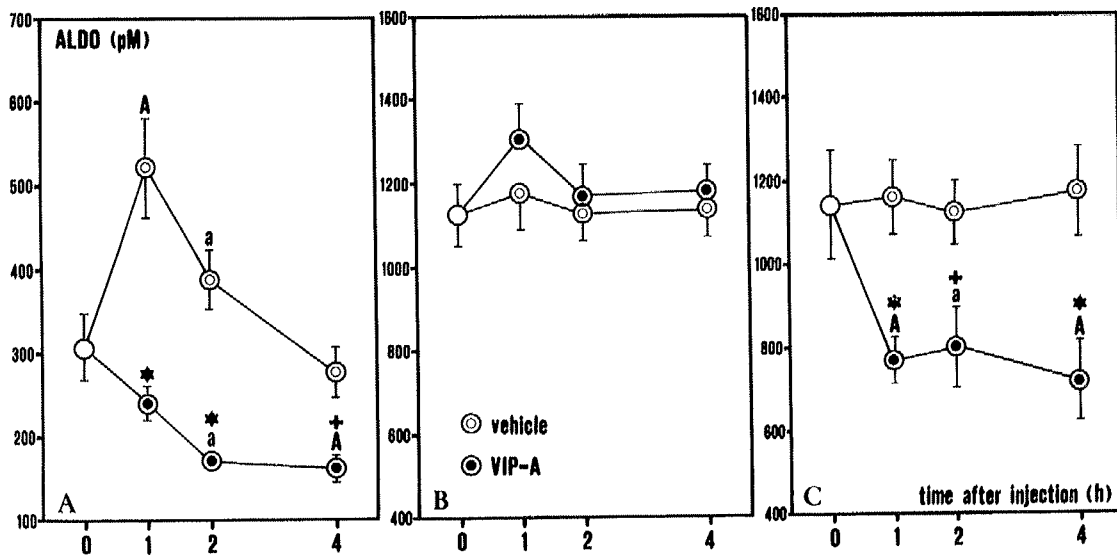


Fig. 2 Effect of VIP-A on ALDO plasma concentration of normal (A), ether-stressed (B) and cold-stressed (C) rats. Data are means \pm SE ($n = 6$). $^aP < 0.05$ and $^AP < 0.01$ from 0-h group; $^+P < 0.05$ and $^*P < 0.01$ from the respective control (vehicle-injected) group.

10%). These raises lasted without significant changes until 4 h, and were not modified by the vehicle injection (Figs 1-4).

VIP-A did not affect ACTH and ALDO responses to ether stress, though small non-significant increases were observed at 1 h (Figs 1 and

2). B response to ether stress was not significantly modified at 1 h and 2 h, but it underwent to a slight potentiation (46%) at 4 h (Fig. 3). Conversely, VIP-A administration strikingly depressed ACTH and ALDO responses to cold stress at 1, 2 and 4 h (about -32/-42%, and -30/-40%, respectively), and

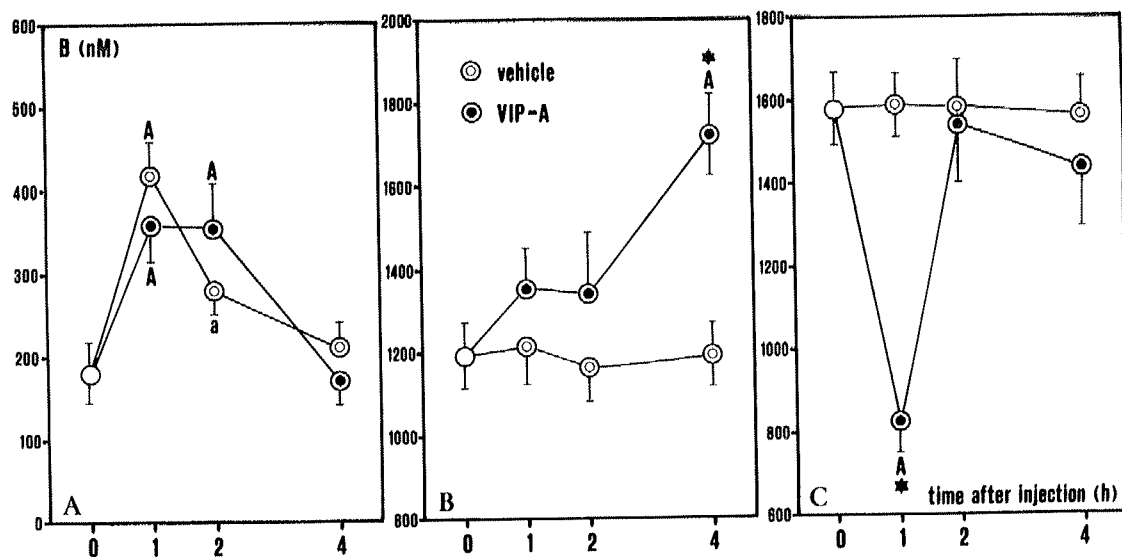


Fig. 3 Effect of VIP-A on B plasma concentration of normal (A), ether-stressed (B) and cold-stressed (C) rats. Data are means \pm SE (n = 6). *P < 0.05 and ^AP < 0.01 from 0-h group; *P < 0.01 from the respective control (vehicle-injected) group.

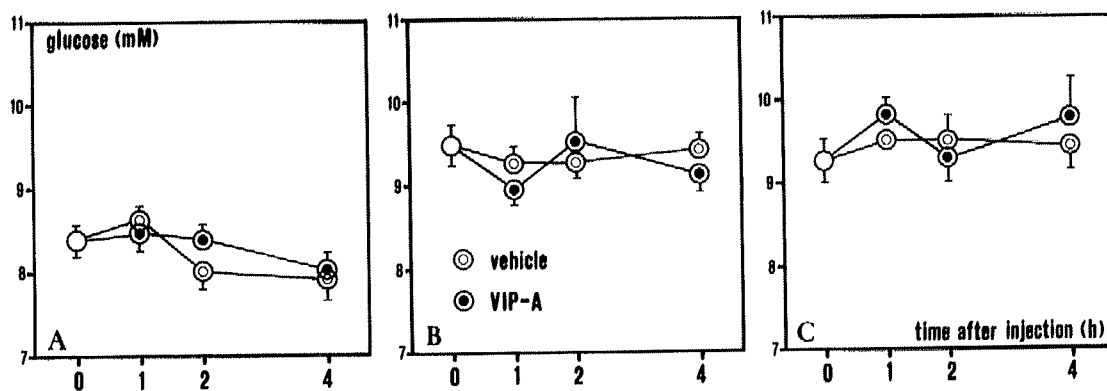


Fig. 4 Effect of VIP-A on glycemia of normal (A), ether-stressed (B) and cold-stressed (C) rats. Data are means \pm SE (n = 6).

that of B at 1 h (-48%) (Figs 1-3). The increase in glycemia induced by either kinds of stress was not modified by VIP-A (Fig. 4).

Discussion

The effects of VIP on the secretion of ACTH and glucocorticoid hormones were the object of several investigations, whose results are not always in complete agreement.

Evidence indicates that VIP stimulates ACTH secretion of the atT20 clonal strain of mouse pitu-

itary tumor cells and human pituitary adenoma cells,²²⁻²⁴ but not of normal corticotropes.²⁵⁻³⁰ However, VIP potentiates CRH-induced ACTH release by superfused rat anterior-pituitary fragments.³⁰ Contrasting results are reported in vivo: Nussdorfer and Mazzocchi³¹ did not find any effect of VIP on the ACTH plasma concentration of dexamethasone-suppressed rats, while Pralong et al³² reported a 10-fold rise in ACTH blood level in rats after 30 min of intravenous VIP infusion. VIP was reported to enhance glucocorticoid secretion in hypophysectomized dogs³³ and intravenously

infused rats,³² but not in dexamethasone-suppressed rats;³¹ conversely a prolonged VIP administration appears to lower plasma B concentration in normal rats and to decrease B output by adrenal slices.^{19,34} A direct effect of VIP on inner adrenocortical cells has been shown, but it seems to be due to the agonistic interaction of VIP with ACTH receptors.^{35,36}

Our present results showing that VIP-A does not significantly affect the transient rises in ACTH and B plasma levels induced by the vehicle injection, cast doubts about the involvement of endogenous VIP in the physiological regulation of basal HPA-axis function in rats. However, they clearly show that VIP-A suppresses ACTH and B responses to cold stress, without notably affecting those to ether stress.

These last findings indicate that different mechanisms underly the effects of the two types of stress, and strongly suggest that endogenous VIP plays a pivotal role in the mediation of cold-induced activation of rat HPA axis. It may be tentatively suggested that cold-evoked signals, but not ether inhalation-evoked ones, acting at the hypothalamic levels induce the release into the portal circulation (see Introduction) of VIP, that stimulates pituitary corticotropes to secrete ACTH, which in turn enhances glucocorticoid secretion. Our data also rule out the possibility that changes in blood glucose concentration may participate in the cold stress-induced activation of HPA axis mediated by endogenous VIP.

Rather intriguing results were obtained as to the effects of VIP-A on plasma ALDO concentration: in fact, VIP-A not only depresses ALDO response to cold-stress, a finding in keeping with the above discussed ones, but also markedly lower basal plasma concentration of ALDO. Two lines of evidence may explain this last result: (i) adrenal glucocorticoid secretion is almost exclusively ACTH dependent, while mineralocorticoid secretion undergoes a complex multifactorial control;³⁷⁻³⁹ and (ii) VIP exerts a direct action on rat-adrenal zona glomerulosa. To summarize: rat zona-glomerulosa cells possess specific VIP receptors,⁴⁰ and VIP exerts a strong ALDO secretagogue action on perfused rat capsule-zona glomerulosa preparations,⁴¹ adrenal perfused *in situ*^{42,43} and adrenal slices.⁴⁴ Thus, our findings may be inter-

preted as indicating that, under basal conditions, endogenous VIP exerts a tonic direct stimulatory action on ALDO secretion of rat zona glomerulosa.

In conclusion, the use of a specific VIP-receptor antagonist has provided proofs that endogenous VIP is involved in the regulation of basal mineralocorticoid secretion in rats, and that a VIP-ergic mechanism plays an essential role in the cold stress-induced activation of HPA axis in this species.

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References

1. Loren, L., Emson, P. C., Fahrenkrug, J. et al. Distribution of vasoactive intestinal polypeptide in the rat and mouse brain. *Neuroscience* 1979; 4: 1953-1976.
2. Lam, K. S. L. Vasoactive intestinal peptide in the hypothalamus and pituitary. *Neuroendocrinology* 1991; 53 (Suppl. 1): 45-51.
3. Malendowicz, L. K. Involvement of neuropeptides in the regulation of growth, structure and function of the adrenal cortex. *Histol. Histopathol.* 1993; 8: 173-186.
4. Toni, R., Kakucska, I., Mosca, S., Marrama, P. and Lechan, R. M. Hypothyroidism increases vasoactive intestinal polypeptide (VIP) immunoreactivity and gene expression in the rat hypothalamic paraventricular nucleus. *Endocrinology* 1982; 131: 976-978.
5. Mezey, E. and Kiss, J. Z. Vasoactive intestinal peptide-containing neurons in the paraventricular nucleus may participate in regulating prolactin secretion. *Proc. Nat. Acad. Sci. USA* 1985; 82: 245-247.
6. Ceccatelli, S., Eriksson, M. and Hokfelt, T. Distribution and coexistence of corticotropin-releasing factor-, neurotensin-, enkephalin-, cholecystokinin-, galanin-, and vasoactive intestinal polypeptide/peptide histidine isoleucine-like peptides in the parvocellular part of the paraventricular nucleus. *Neuroendocrinology* 1989; 49: 309-323.
7. Samson, W. K., Said, S. I., Graham, J. W. and McCann, S. M. Vasoactive intestinal polypeptide concentrations in median eminence of hypothalamus. *Lancet* 1978; II: 901-902.
8. Said, S. I. and Porter, J. C. Vasoactive intestinal polypeptide: release into hypophyseal portal blood. *Life Sci.* 1979; 24: 227-230.
9. Morel, G., Besson, J., Rosselin, G. and Dubois, P. M. Ultrastructural evidence for endogenous vasoactive intestinal peptide-like immunoreactivity in the pituitary gland. *Neuroendocrinology* 1982; 34: 85-89.
10. Arnaout, M. A., Garhwaite, T. L., Martinson, D. R. and Hagen T. C. Vasoactive intestinal polypeptide is synthesized

- in anterior pituitary tissues. *Endocrinology* 1986; 119: 2052–2057.
11. Calvo, J. J., de Carvalho, L. F., Gonzales, R., Burnet, P. W. J., Ghatei, M. A. and Bloom, S. R. Effect of ACTH on VIP and galanin release from the pituitary. *Endocrinology* 1990; 126: 1283–1287.
 12. Chew, L. J., Murphy, D. and Carter, D. A. Differential use of 3' poly(A) addition sites in vasoactive intestinal peptide messenger ribonucleic acid of the rat anterior pituitary gland. *J. Neuroendocrinol.* 1991; 3: 351–355.
 13. Bryant, M., Bloom, S. R., Polak, J. M., Albuquerque, R. H., Modlin, I. and Pearse, A. G. Possible dual role for vasoactive intestinal peptide as gastro-intestinal hormone and neurotransmitter substance. *Lancet* 1976; 1: 991–993.
 14. Linnoila, R. I., Diaugustine, R. P., Hervonen, A. and Miller, R. J. Distribution of (Met⁵)- and (Leu⁵)-enkephalin-, vasoactive intestinal polypeptide- and substance P-like immunoreactivities in human adrenal glands. *Neuroscience* 1980; 5: 2247–2259.
 15. Hokfelt, T., Lundberg, J. M., Schultzberg, M. and Fahrenkrug, J. Immunohistochemical evidence for a local VIP-ergic neuron system in the adrenal gland of the rat. *Acta Physiol. Scand.* 1981; 113: 575–576.
 16. Holzwarth, M. A. The distribution of vasoactive intestinal peptide in the rat adrenal cortex and medulla. *J. Auton. Nerv. Syst.* 1984; 11: 269–283.
 17. Kondo, H., Kuramoto, H. and Fujita, T. An immunoelectron-microscopic study of the localization of VIP-like immunoreactivity in the adrenal gland of the rat. *Cell Tissue Res.* 1986; 245: 531–538.
 18. Maubert, E., Tramu, G., Croise, D., Beauvillain, C. J. and Dupouy, J. P. Co-localization of vasoactive intestinal polypeptide in nerve fibers of the rat adrenal gland. *Neurosci. Lett.* 1990; 113: 121–126.
 19. Malendowicz, L. K. and Nussdorfer, G. G. Unusual effect of prolonged vasoactive intestinal peptide (VIP) administration on the adrenal growth and corticosterone secretion in the rat. *Neuropeptides* 1993; 25: 145–150.
 20. Waelbroeck, M., Robberecht, P., Coy, D. H., Camus, J. C., DeNeef, P. and Christophe, J. Interaction of growth hormone-releasing factor (GRF) and 14 GRF analogs with vasoactive intestinal peptide (VIP) receptors of rat pancreas. Discovery of (N-Ac-Tyr¹,D-Phe²)-GRF-(1-29)NH₂ as VIP antagonist. *Endocrinology* 1985; 116: 2643–2649.
 21. Lesniewska, B., Miskowiak, B., Nowak, M. and Malendowicz, L. K. Sex differences in adrenocortical structure and function. XXVII. The effect of ether stress on ACTH and corticosterone in intact, gonadectomized, and testosterone- or estradiol-replaced rats. *Res. Exp. Med.* 1990; 190: 95–103.
 22. Oliva, D., Nicosia, S., Spada, A. and Giannattasio, G. VIP stimulates ACTH release and adenylate cyclase in human ACTH-secreting pituitary adenomas. *Eur. J. Pharmacol.* 1982; 83: 101–105.
 23. Reisine, T., Heisler, S., Hook, V. Y. and Axelrod, H. Multireceptor induced release of adrenocorticotropin from anterior pituitary tumor cells. *Biochem. Biophys. Res. Commun.* 1982; 108: 1251–1255.
 24. Westendorf, J. M. and Schonbrunn, A. Peptide specificity for stimulation of corticotropin secretion: activation of overlapping pathways by the vasoactive intestinal peptide family and corticotropin-releasing factor. *Endocrinology* 1985; 116: 2528–2535.
 25. Rotsztejn, W. H., Benoist, L., Besson, J. et al. Effect of vasoactive intestinal peptide (VIP) on the release of adeno- hypophysial hormones from purified cells obtained by unit gravity sedimentation. *Neuroendocrinology* 1980; 31: 282–286.
 26. Westendorf, J. M., Phillips, M. A. and Schonbrunn, A. Vasoactive intestinal peptide stimulates hormone release from corticotropic cells in culture. *Endocrinology* 1983; 112: 550–557.
 27. Tilders, F., Tatemoto, K. and Berkenbosch, F. The intestinal peptide PHI-27 potentiates the action of corticotropin-releasing factor on ACTH from rat pituitary fragments in vitro. *Endocrinology* 1984; 115: 1633–1635.
 28. Nicholson, S. A., Adrian, T. E., Gillhann, B., Jones, M. T. and Bloom, S. R. Effect of hypothalamic neuropeptides on corticotrophin release from quarters of rat anterior pituitary gland in vitro. *J. Endocrinol.* 1984; 100: 219–226.
 29. Vigh, S. and Schally, A. V. Interactions between hypothalamic peptides in a superfused pituitary cell system. *Peptides* 1984; 5: 241–247.
 30. Leonard, J. F., Bluet-Pajot, M. T., Oliver, C. and Cordon, C. Interaction of vasoactive intestinal peptide (VIP) and growth hormone releasing factor (GRF) with corticotropin releasing factor (CRF) on corticotropin secretion in vitro. *Neuropeptides* 1988; 12: 131–133.
 31. Mazzocchi, G., Robba, C., Malendowicz, L. K. and Nussdorfer, G. G. Stimulatory effect of vasoactive intestinal peptide (VIP) on the growth and steroidogenic capacity of rat adrenal zona glomerulosa. *Biomed. Res.* 1987; 8: 19–23.
 32. Pralong, F. P., Corder, R. and Gaillard, R. C. Responses of the rat pituitary-adrenal axis to hypotensive infusions of corticotropin-releasing factor, vasoactive intestinal peptide and other depressor agents. *Regul. Pept.* 1991; 32: 217–226.
 33. Bloom, S. R., Edwards, A. V. and Jones, C. T. Adrenal cortical responses to vasoactive intestinal peptide in conscious hypophysectomized calves. *J. Physiol.* 1987; 391: 441–450.
 34. Nussdorfer, G. G. and Mazzocchi, G. Vasoactive intestinal peptide (VIP) stimulates aldosterone secretion by rat adrenal glands in vivo. *J. Steroid Biochem.* 1987; 26: 203–205.
 35. Li, Z. G., Queen, G. and Labelle, F. S. Adrenocorticotropin, vasoactive intestinal polypeptide, growth hormone-releasing factor, and dynorphin compete for common receptors in brain and adrenal. *Endocrinology* 1990; 126: 1327–1333.
 36. Mazzocchi, G., Malendowicz, L. K. and Nussdorfer, G. G. Stimulatory effect of vasoactive intestinal peptide (VIP) on the secretory activity of dispersed rat adrenocortical cells. Evidence for the interaction of VIP with ACTH receptors. *J. Steroid Biochem. Mol. Biol.* 1994; 48, in press.
 37. Nussdorfer, G. G. Cytophysiology of the adrenal cortex. *Int. Rev. Cytol.* 1986; 98: 1–405.
 38. Quinn, S. J. and Williams, G. H. Regulation of aldosterone secretion. In: James, V. H. T. (ed.) *The Adrenal Gland*. New York: Raven Press, 1992, 159–189.
 39. Vinson, G. P., Whitehouse, B. J. and Hinson, J. P. *The Adrenal Cortex*. Englewood Cliffs: Prentice Hall, 1992.
 40. Cunningham, L. A. and Holzwarth, M. A. Autoradiographic distribution of ¹²⁵I-VIP binding in the rat adrenal cortex. *Peptides* 1989; 10: 1105–1108.
 41. Cunningham, L. A. and Holzwarth, M. A. Vasoactive intestinal peptide stimulates adrenal aldosterone and corticosterone secretion. *Endocrinology* 1988; 122: 2090–2097.
 42. Ehrhart-Bornstein, M., Bornstein, S. R., Scherbaum, W. A., Pfeiffer, E. F. and Holst, J. J. Role of vasoactive intestinal peptide in a neuroendocrine regulation of the adrenal cortex. *Neuroendocrinology* 1991; 54: 623–628.
 43. Hinson, J. P., Kapas, S., Orford, C. D. and Vinson, G.

- P. Vasoactive intestinal peptide stimulation of aldosterone secretion by the rat adrenal cortex may be mediated by the local release of catecholamines. *J. Endocrinol.* 1992; 133: 253-258.
44. Mazzocchi, G., Malendowicz, L. K., Meneghelli, V., Gottardo, G. and Nussdorfer, G. G. Vasoactive intestinal polypeptide (VIP) stimulates hormonal secretion of the rat adrenal cortex in vitro: evidence that adrenal chromaffin cells are involved in the mediation of the mineralocorticoid, but not glucocorticoid secretagogue action of VIP. *Biomed. Res.* 1993; 14: 435-440.